PK analysis were collected for 24 h following each dose of simvastatin. Plasma samples were assayed for dasatinib, simvastatin and simvastatin acid by validated LC MS/MS. PK parameters were derived from plasma concentration versus time data by noncompartmental methods.

Results: Dasatinib increased the C_{max} of simvastatin by 37% and AUC_{∞} by 20% versus simvastatin alone. Dasatinib also increased the C_{max} of simvastatin acid by 41% and AUC_{∞} by 27% versus simvastatin alone.

Table: Summary statistics for simvastatin pharmacokinetic parameters

Treatment	C _{max} (ng/mL) Geometric mean (CV%)	AUC _∞ (ng⋅h/mL) Geometric mean (CV%)	AUC _{0-T} (ng·h/mL) Geometric mean (CV%)	T _{max} (h) Median (min, max)	t _{1/2} (h) Mean (SD)
Simvastatin 80 mg (n = 48) Simvastatin 80 mg and dasatinib 100 mg (n = 48)	26.68 (57) 36.53 (57)	117.95 (80) 141.29 (68)	108.05 (74) 132.97 (66)	1.50 (0.50, 8.00) 1.00 (1.00, 5.00)	6.65 (3.00) 5.16 (2.85)

 C_{max} = maximum plasma concentration; AUC = area under plasma concentration—time curve; T_{max} = time to maximum plasma concentration; $t_{1/2}$ = terminal half-life; SD = standard deviation; CV% = coefficient of variation.

Conclusions: Dasatinib increases exposure to the CYP3A4 substrates simvastatin and simvastatin acid. Due to the small effect size, these findings are not felt to be clinically significant.

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Delivery of indenoisoquinoline using customized releasable PEG linkers

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Background: Indenoisoquinolines are novel topoisomerase I inhibitors with good *in vitro* anti-tumor efficacies but suffer from poor water solubility. Using customized releasable PEG linker technology, we have successfully solublized the lead indenoisoquinoline compound, MJ-III-65 (NSC 706744), to facilitate its administration to animals.

Material and Methods: NSC 706744 and customized releasable PEG linkers were synthesized separately according to previously published methods. The selected PEG linker, PEG-RNL9, was conjugated with NSC 706744 through its secondary amine group. Two different molecular weights of PEG were used, i.e. 20k PEG to give EZN-2087 (NSC 735982) and 40k PEG to give EZN-2088 (NSC 735983). The NSC 735982 was determined to contain 4.4% NSC 706744 by weight while the NSC 735983 was 3% by weight. The *in vivo* hollow fiber assay (HFA) in mice was conducted per previously published methods using NSC 735982 at equivalent active doses of 12 and 18 mg/kg/dose and NSC 735983 at equivalent active doses of 9 and 12 mg/kg/dose.

Results: The PEG conjugates were stable in saline at room temperature for at least 4 hours and the half-lives in rat plasma were about 4 hours. This feature enabled administration of PEG conjugates of NSC 706744 in vivo. The in vivo studies of both PEGylated compounds, particularly NSC 735983 showed antitumor activity. Using the published scoring comparison, NSC 735982 produced scores of 12/48 IP and 4/48 SC for a total score of 16/96. Of greater note, the doses of NSC 735983 tested resulted in scores of 28/48 IP and 10/48 SC. The NSC 735983 total score of 38/96 places it in the top 3% of the 3604 compounds evaluated in the hollow fiber assay to date.

Conclusions: PEGylation of indenoisoquinoline compound using customized releasable PEG linkers has successfully solubilized the lead compound NSC 706744. This feature enabled *in vivo* evaluation of NSC 706744 in a form that was potentially more bioavailable than the parent compound. The NCI's *in vivo* HFA study revealed that anti-tumor efficacy could be achieved through this modification. In the future, customized releasable PEG linkers with different half-lives can be applied to this compound to further study the relationship between pharmacokinetic profile and efficacy.

POSTER

Enhanced antitumor activity and safety of albumin-bound nab-docetaxel versus polysorbate 80-based docetaxel

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Background: Docetaxel (Taxotere®; TAX) is currently formulated in the solvent polysorbate 80. Removal of solvents from taxane formulations, as in the case of albumin-bound *nab*-paclitaxel (Abraxane®), has resulted in significantly higher response rates and greater safety [Gradishar, JCO 2005;23:7794]. Polysorbate 80 strongly inhibited the binding of taxanes to albumin, possibly inhibiting albumin-based drug transport through the gp60 endothelial receptor [Desai, EORTC-NCI-AACR, 2004] and consequently reducing albumin-binding to tumor-secreted protein (SPARC). We compared the efficacy, toxicity, and pharmacokinetics (PK) of TAX and solvent-free *nab*-docetaxel.

Methods: *nab*-Docetaxel and TAX were tested in nude mice (q4dx3) in HCT-116 human colon carcinoma xenograft (equitoxic doses of 22 and 15 mg/kg, respectively;10/group) and PC3 human prostate xenograft (0, 10, 15, 20, or 30 mg/kg *nab*-docetaxel or 10 mg/kg TAX; 6/group). *nab*-Docetaxel was compared with TAX in rats for single-dose toxicity (25, 50, 75, 100, and 125 mg/kg), multiple-dose toxicity (5, 10, 15, 30, and 50 mg/kg q4dx3), and PK (10, 20, and 30 mg/kg) (all 3/group).

Results: Both drugs were effective in HCT-116 xenograft; at equitoxic doses, nab-docetaxel exhibited greater antitumor activity than TAX (P<0.0001, ANOVA). In PC3 xenograft, TAX was toxic (6/6 rats died); nab-docetaxel was well tolerated at all doses (1 death [15 mg/kg]). Tumor suppression was observed at all nab-docetaxel doses (6/6 complete regressions at 30 mg/kg). In the single-dose study, mortality was more rapid and complete for TAX than for nab-docetaxel at all doses. LD $_{50}$ was 63 mg/kg for nab-docetaxel and ~12.5 mg/kg for TAX. In the multiple-dose study, mortality was similar for both drugs, with complete survival only at the lowest dose (5 mg/kg), where weight loss, neutropenia, and organ toxicity were substantially less for nab-docetaxel than for TAX. PK was similar for nab-docetaxel and TAX at 10 mg/kg; however, at 20 and 30 mg/kg, C_{max} and AUC were lower and V_z and V_{ss} were higher for nab-docetaxel and exponential for TAX.

Conclusions: nab-Docetaxel was less toxic than TAX. nab-Docetaxel showed greater antitumor activity than TAX against HCT-116 colon and PC3 prostate tumors. PK results suggest solvent-mediated sequestration of docetaxel in plasma for TAX. These observations are similar to those seen for nab-paclitaxel vs solvent-based paclitaxel.

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Pharmacokinetics of IHL-305, a novel PEGylated liposome containing irinotecan, in rats and dogs

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Background: Recently, liposomal formulations of anticancer drugs have been developed to enhance their pharmacologic activity and/or to reduce their toxicity. IHL-305 is a PEGylated liposome containing irinotecan. In human xenografts, IHL-305 showed superior antitumor activity to irinotecan hydrochloride (CPT-11). We compared the plasma pharmacokinetics and excretion after intravenous administration of IHL-305 with those of CPT-11 in rats and dogs.

Materials and Methods: IHL-305 or CPT-11 was injected intravenously (i.v.) to SD rats (3, 10, and 30 mg/kg), and to beagle dogs (1, 3, and 10 mg/kg) in plasma pharmacokinetic studies. Plasma concentrations of irinotecan and its metabolites were analyzed by fluorescence-HPLC. In excretion studies, IHL-305 containing [¹⁴C]irinotecan or [¹⁴C]CPT-11 was given i.v. (10 mg/kg) to male rats and a dog. Urine, feces, and bile (rats only) were collected and radioactivity (RA) in excreta was analyzed.

Results: Irinotecan decreased monoexponentially with almost linear pharmacokinetics in both animals after IHL-305 dosing at the doses examined. The total clearances of irinotecan after IHL-305 dosing were about 1/500 and 1/100 of those after CPT-11 dosing in rats and dogs, respectively. The distribution volume was about 1/80 to 1/30 of those after CPT-11 dosing. No apparent gender difference was observed in rats. The AUC ratios of lactone to carboxylate forms of irinotecan were about 150 and 1.6 after IHL-305 and CPT-11 dosing in dogs, respectively. IHL-305 increased the AUC and mean residence time of 7-ethyl-10-hydroxycamptothecin (SN-38), an active metabolite of irinotecan, by about 3- and 9-fold compared with those after CPT-11 dosing in rats, respectively. Urinary and fecal excretion of RA after IHL-305 dosing was almost completed at 48 hours, whereas at 24 hours after CPT-11 dosing in rats.